

Procedure for Handling Positive Blood Cultures from Patients with Suspect or Confirmed EVD/VHF

Refer to IPAC website, <http://ipac.vch.ca/Pages/Emerging-Issues.aspx>, to ensure you are using the most updated protocol.

VGH Medical Microbiology Laboratory Guidelines
Positive blood cultures from patients with suspect Ebola Virus Disease (EVD) or
other Viral Hemorrhagic Fevers (VHF)

Blood Culture Technologist:

v		Step
	1.	BACTEC FX signals positive blood culture.
	2.	Door of BACTEC has pink "Suspect Ebola VD/VHF" sticker. Open door to see which bottle is positive. Positive blood culture bottle observed to have pink Ebola VD/VHF sticker. DO NOT REMOVE BOTTLES. Shut door.
	3.	Notify lab supervisor, medical microbiologist on call, and technologist in AFB room. Discuss patient's current status with medical microbiologist before proceeding. If patient's status is still pending or if they have tested positive for Ebola VD/VHF proceed as described.
	4.	The TB technologist will process the positive blood cultures.
	5.	The microbiology supervisor will designate a second laboratory technologist to assist.
	6.	If a blood culture becomes positive off-shift, technologist trained in Ebola/VHF procedures/VHF will need to be called in. Consult with medical microbiologist. An on call list of trained technologists is provided on the huddle board.
	7.	Full PPE will be worn by micro technologist when processing positive blood culture. Follow the donning and doffing protocol check list: <i>'Donning and Doffing Viral Hemorrhagic Fever (VHF) Specimen Processing PPE Check List for VGH Laboratory Technologists'</i> .
	8.	The processing of the positive blood culture is to be performed by one technologist, while the other assists by reading and recording the process from checklist.

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AFB technologist:

v		Step
	1.	Suspend processing TB samples at an appropriate point.
	2.	Remove all biohazardous waste from the AFB Room. Tie off all bags, label with "autoclave before disposal" sticker and place in wash up area for autoclaving. It is imperative that all biohazard waste is removed before proceeding.
	3.	Remove all items from inside the BSC
	4.	Clear and de-clutter work area outside the hood
	5.	Begin to prepare the AFB room.

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A: AFB Room Check list

*Refer to 'AFB Room Supply Lists – Suspect EVD/VHF Specimen Processing' document on IPAC website, <http://ipac.vch.ca/Pages/Emerging-Issues.aspx>, for full AFB Room Supply Lists

v		Step
	1.	Turn on the BSC if not already running. Note time on a post it note and attach to outside of BSC. Ensure it runs at least 30 minutes before use.
	2.	Prepare a working area inside the hood by placing down two blue absorbent pads.
	3.	Ensure gas is turned off in the BSC. Place the 95°C heating block in the BSC, turn on, checking temperature setting, and make sure no plastics or flammables are close by.
	4.	Ensure all the required supplies are ready for use in 'Ebola VD/VHF Positive Blood Culture' designated bin, following the list provided on the 'AFB Room Supplies List - Suspect EVD/VHF'. Place bin on the bench to the right of the BSC.
	5.	Clear the top of the cart to be used for the transfer of the blood culture bottles to the BSC from the BACTEC Instrument.
	6.	Place inside the BSC: <ul style="list-style-type: none"> • Accel Five TB Disinfectant • Accel INTERvention disinfectant wipes - ready to be dispensed. • Alcohol based hand rub (ABHR)
	7.	Prepare Coplin jar with enough fresh methanol to cover the entire surface of a slide and place in BSC.
	8.	Place all required supplies in the BSC: media, anaerobic jar/indicator (for B bottles only), pack of plastic loops, venting units, small red sharps container, frosted slides, slide case, pencil, sharpie, biohazard Ziploc bags and double bagged waxed paper waste bucket.
	9.	Ensure 2 copies of checklist: 'Donning and Doffing Viral Hemorrhagic Fever (VHF) Specimen Processing PPE Check List - for VGH Laboratory Technologists', one for each technologist, are brought into the AFB room.
	10.	Ensure proper airflow by maintaining adequate space between all objects inside the cabinet and keeping the front grill clear. Ensure proper air flow according to safe practice use of a BSC (Refer to instructions for proper use of BSC – laminated copy posted near AFB BSC)

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	11.	<p>Place blue absorbent mat on top of the cart and add the following supplies to the mat:</p> <ul style="list-style-type: none"> • Accel Five TB disinfectant • Accel INTERvention disinfectant wipes - ready to be dispensed. • Alcohol based hand rub (ABHR) • A hard plastic TDG container (with absorbent material at the bottom) and lid
	12.	<p>Place blue biohazardous waste bin and lid for EVD/VHF waste inside AFB room along with all the required bin labels. (One blue bin is stored in the wash up room 1351 and one in the TDG area in microbiology lab). Ensure that the absorbent material provided with the bin is placed at the bottom of the bin and that bin is double lined with the red biohazard bags provided.</p>
	13.	<p>Place sign on the door "Ebola VD/VHF Containment/Processing DO NOT ENTER".</p>

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B: Donning of PPE:

v	Step
1.	Follow the lab protocol for donning using the PPE checklist: <i>'Donning and Doffing Viral Hemorrhagic Fever (VHF) Specimen Processing PPE Check List for VGH Laboratory Technologists'</i>
2.	All personnel must be trained in PPE protocol and be fit tested for N95 within the last year.
3.	Designated donning area is outside AFB Room, Room 1105.
4.	Donning will not commence until both technologists are present.
5.	Both technologists must sign the <i>'Specimen Handling Contact List'</i> which is located in the TDG Area.
6.	Both technologists are to be engaged and assist one another, one donning at a time while the other instructs, observes and records the process.

C: Subculture of Positive BACTEC Bottles:

- Note: Accel INTERvention disinfecting wipes require **one minute** contact time
Accel 5 TB disinfectant (liquid) requires **five minute** contact time
- Accel INTERvention disinfecting wipes = 'disinfectant wipes'
- Accel 5 TB disinfectant (liquid) = 'disinfectant (liquid)'
- Alcohol based hand rub = 'ABHR'
- Blue biohazardous waste container = 'blue bin'

v	Step
1.	Wearing full PPE , take the prepared cart carrying the TDG container to the BACTEC Instrument, remove positive blood culture(s) from BACTEC and place in TDG container. Close lid on the TDG container.
2.	Disinfect outer gloves using disinfectant wipe or alcohol based hand rub (ABHR). Allow to dry.
3.	Confirm that AFB room is negative pressure prior to use by taking a tissue and placing it under the gap under the door. Tissue should move with the air pressure in an inward direction towards the inside of room.
4.	Using the cart, take TDG container into AFB room, close door and place in BSC.
5.	Remove positive blood culture bottles and wipe top of bottles with disinfectant wipes.
6.	Clean outer gloves using disinfectant wipe or ABHR. Allow to dry.

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7.	Label slides and all plates with specimen number, patient name, bottle type and date.
8.	Vent bottles with venting unit. DO NOT RECAP AFTER USE.
9.	Inoculate BAP, MRSS, Chocolate, MacConkey, CNA and, if a B bottle, a Brucella plate.
10.	Make smear. Allow to air dry.
11.	Using a disposable plastic loop, gently spread out inoculums for isolation being careful not to splash.
12.	Clean outer gloves using disinfectant wipe or ABHR. Allow to dry.
13.	As soon as slide is dry, put slide in Coplin jar with methanol, making sure entire slide is covered. Set timer for 30 minutes.

14.	Disinfect outer gloves using disinfectant wipe or ABHR. Allow to dry.
15.	Wipe outer surfaces of all plates with fresh disinfectant wipes, allowing one minute contact time.
16.	Seal all plates with Parafilm, add pink Ebola VD/VHF sticker to each plate, and place aerobic plates into a biohazard bag. Wipe surface of Biohazard bag with disinfectant wipe. Allow to dry and label with an Ebola VD/VHF sticker.
17.	Place plates into a rack. Apply an Ebola VD/VHF sticker to the rack so that it is clearly visible.
18.	Place the anaerobic plates into an anaerobic jar and set the jar up.
19.	Wipe the surface of the jar with disinfectant wipes.
20.	Disinfect outer gloves using disinfectant wipe or ABHR. Allow to dry.
21.	Incubate primary plates and anaerobic jar in an assigned area in the CO2 incubator in AFB room.
22.	Carefully remove venting unit from blood culture bottle and place in sharps container.
23.	Wipe top of bottles with disinfectant wipes. Allow to dry.
24.	Disinfect outer gloves using disinfectant wipes or ABHR. Allow to dry.
25.	Place blood culture bottles into biohazard bags and place in new TDG container. Seal container and wipe outer surface with disinfectant wipe. Allow to dry.
26.	Label top and sides of TDG container with the accession number and Ebola VD/VHF stickers and store in the BSC until blood culture is finalized.
27.	After 30 minutes in methanol, remove slide and allow to air dry.
28.	Ensure that the heating block has come to temperature and place slide on 95°C heating block to inactivate the virus. Set timer for one hour.

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	29.	Wipe the surface of the Accel INTERVention wipes container with disinfectant wipe and allow to dry.
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D: Doffing of Personnel Protective Equipment

The procedure for removing protective equipment is critical and strict adherence to the order of removal of PPE must be followed.

Follow VGH Lab Doffing Protocol using the PPE checklist: **'Donning and Doffing Viral Hemorrhagic Fever (VHF) Specimen Processing PPE Check List for VGH Laboratory Technologists'**. (Refer to IPAC website, <http://ipac.vch.ca/Pages/Emerging-Issues.aspx>, to ensure you are using the most updated protocol.)

Do not rush!

The AFB room is the designated doffing area

- Both technologists are to be engaged and assist one another in the doffing process, each taking turns doffing while the other instructs, observes, assists and records the process, ensuring each step is followed
- Note: Accel INTERVention disinfecting wipes require one minute contact time
Accel 5 TB disinfectant (liquid) requires five minute contact time
 - Accel INTERVention disinfecting wipes = 'disinfectant wipes'
 - Accel 5 TB disinfectant (liquid) = 'disinfectant (liquid)'
 - Alcohol based hand rub = 'ABHR'
 - Blue biohazardous waste container = 'blue bin'

v		Step
	1.	All PPE should be removed in the AFB room and should be discarded into the blue bin, double-lined with red biohazard bags (do NOT compress). Follow checklist using buddy system until completion of PPE Doffing.
	2.	Following full doffing procedure, disinfect hands using ABHR, allow to dry.
	3.	Leave AFB Room, close door and secure room with sign "Suspect Ebola VD/VHF Sample Containment / Processing DO NOT ENTER" on door and place barrier/masking tape over door to prevent accidental entry.
	4.	AFB room cannot be used for routine purposes until room cleaning is completed and all Ebola VD/VHF waste is removed.

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E: Inactivated Blood Culture Slide:

v		Step
	1.	Donning full PPE following: <i>'Donning and Doffing Viral Hemorrhagic Fever (VHF) Specimen Processing PPE Check List for VGH Laboratory Technologists'</i> , the TB tech will prepare to independently re-enter the AFB room after the slide inactivation is complete (one hour at 95 °C). Have the buddy technologist or a Trained Observer assist with donning procedure in donning area outside AFB Room.
	2.	Enter AFB Room. Unplug heating block.
	3.	Place inactivated slide into slide case and label case with 'Inactivated' sticker.
	4.	Call or alert blood culture tech (or evening tech) to stand outside AFB room to receive slide from TB Tech for staining. TB Tech will open AFB room door and pass the slide through door directly to blood culture tech. Blood culture tech will notify medical microbiologist on call to review gram stain and will call the results to the appropriate location. Medical microbiologist on call will discuss with physician per clinical judgement.
	5.	TB tech will re-enter AFB room to perform room clean up.

F: AFB Room Interim Cleanup by TB Technologist:

- Note: Accel INTERvention disinfecting wipes require **one minute** contact time
Accel 5 TB disinfectant (liquid) requires **five minute** contact time
- Accel INTERvention disinfecting wipes = 'disinfectant wipes'
- Accel 5 TB disinfectant (liquid) = 'disinfectant (liquid)'
- Alcohol based hand rub = 'ABHR'
- Blue biohazardous waste container = 'blue bin'

v		Step
	1.	Wearing full PPE, place all disposable waste, into the blue bin that is double lined with red bags. Do <u>NOT</u> press down to compress.
	2.	Disinfect gloves with disinfectant wipe or Alcohol hand rub. Allow to dry.

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3.	<p>Proceed with a Primary Clean:</p> <p>Wipe all surfaces inside the BSC and all surfaces used outside the BSC using Accel Five TB disinfectant, including the surfaces of containers of any used disinfectants. Using a timer, allow to remain wet 5 minutes and reapply if necessary.</p> <p>Note: The heating block is to remain unplugged inside the BSC until the pm day one work up of blood culture is complete. A clean of the heating block will be done once it has cooled, during the final room clean up.</p>
4.	<p>Perform Secondary Clean by repeating the above cleaning process.</p>
5.	<p>Disinfect gloves with disinfectant wipe or ABHR. Allow to dry.</p>
6.	<p>Wipe the surface of the Accel INTERvention wipes container with disinfectant wipe.</p>
7.	<p>When the primary and secondary cleaning is complete, doff PPE in the AFB Room. Follow the doffing protocol: <i>'Donning and Doffing Viral Hemorrhagic Fever (VHF) Specimen Processing PPE Check List for VGH Laboratory Technologists'</i> with assistance of the buddy technologist or Trained Observer outside AFB room to read checklist, record and observe. Place all PPE into the double-lined blue bin.</p>
8.	<p>Following full doffing procedure, exit room and place sign on the AFB room door: "Suspect Ebola VD/VHF processing/containment DO NOT ENTER". Place barrier/masking tape over the door to prevent accidental entry.</p>

G: PM Growth and Day 1 / Subculture of Primary Plates to Secondary:

- If patient’s status is pending, check with medical microbiologist for any updates before proceeding
- Ask the Medical Microbiologist to sign a ‘Waste Release Form’
- Note: Accel INTERvention disinfecting wipes require **one minute** contact time
Accel 5 TB disinfectant (liquid) requires **five minute** contact time
- Accel INTERvention disinfecting wipes = 'disinfectant wipes'
- Accel 5 TB disinfectant (liquid) = 'disinfectant (liquid)'
- Alcohol based hand rub = 'ABHR'
- Blue biohazardous waste container = 'blue bin'

v		Step
	1	Prepare EVD/VHF Positive Blood Culture Box with all required materials and media plates for subculture of growth to secondary plates. Consider the gram stain results and allow for the possibility of more than one colony type being present. Include disposable loops, blue absorbent pads, a double-lined cardboard bucket for waste, two copies of the donning/doffing protocol and 2 routine lab gowns. Take supplies to AFB Room.
	2.	AFB (or designated) technologist will perform the work up independently but will have a buddy technologist or Trained Observer as assistant for the donning and doffing following the lab PPE donning/doffing check list.
	3.	Sign the ‘Specimen Handling Contact List’. (In TDG area)
	4.	Don PPE with assistant following: ‘Donning and Doffing Viral Hemorrhagic Fever (VHF) Specimen Processing PPE Check List for VGH Laboratory Technologists’.
	5.	Perform primary and secondary clean of previously used heating block using disinfectant wipes and allow to remain wet one minute each time. Remove from BSC.
	6.	Disinfect gloves with disinfectant wipes or alcohol-based hand rub (ABHR). Allow to dry.
	7.	Place blue absorbent pads for a work surface in the BSC.
	8.	Arrange all materials for use inside hood including disinfectant wipes (ready to use) and ABHR. Ensure proper air flow according to safe practice use of a BSC (Refer to instructions for proper use of BSC – laminated copy posted near AFB BSC)

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	9.	Remove primary plates from incubator and place into BSC. Close AFB room door.
	10.	Remove plates from biohazard bag and wipe outside of plates with disinfectant wipes. Allow to dry.
	11.	Read plates; observe growth colony characteristics, consistent with Gram stain?
	12.	If there is no growth, re-wipe entire outside surface of plates with disinfectant wipe, allow to dry. Add Parafilm and place back in biohazard bag. Re-incubate.

	13.	Clean gloves with disinfectant wipes or ABHR. Allow to dry.
	14.	Subculture growth to secondary plates. Pick the top of colony in the last quadrant with growth.
	15.	Do multiple subcultures if different colony types are present. Assistant may record numbered workups if required.
	16.	Disinfect gloves with disinfectant wipes or ABHR. Allow to dry.
	17.	Wipe outside of entire surfaces of secondary plates with disinfectant wipe.
	18.	Seal all plates with Parafilm, place an 'inactivated' label on each plate, place in biohazard bag, add 'inactivated' sticker to bag. Wipe outside of biohazard bag with disinfectant wipes and allow to dry.
	19.	Disinfect gloves with disinfectant wipes or ABHR. Allow to dry.
	20.	Incubate secondary plates in the assigned area of the CO2 incubator in AFB room.
	21.	Replace the primary plates in biohazard bag and wipe bag with disinfectant wipes. Allow to dry.
	22.	Label bag with pink Ebola VD/VHF stickers and keep in the incubator in case needed for further work-up.

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H: AFB Room Clean Up:

- Note: Accel INTERVention disinfecting wipes require **one minute** contact time
Accel 5 TB disinfectant (liquid) requires **five minute** contact time
- Accel INTERVention disinfecting wipes = 'disinfectant wipes'
- Accel 5 TB disinfectant (liquid) = 'disinfectant (liquid)'
- Alcohol based hand rub = 'ABHR'
- Blue biohazardous waste container = 'blue bin'

1.	Wearing full PPE, discard all disposable waste and any unused media into lined blue bin.
2.	Proceed with a Primary Clean: wipe all surfaces inside the BSC and all surfaces used outside the BSC using disinfectant (liquid), including the surface of the disinfectant containers. Note the time and allow to remain wet minimum 5 minutes. Reapply if necessary
3.	Perform a Secondary Clean by repeating the above process.
4.	Wipe all the exterior surfaces of the blue bin with disinfectant wipes. Allow to dry.
5.	Place an absorbent mat on the floor and saturate with disinfectant (liquid). Place the blue bin on top of the mat.

I: Waste removal from AFB room:

- Note: Accel INTERVention disinfecting wipes require **one minute** contact time
Accel 5 TB disinfectant (liquid) requires **five minute** contact time
- Accel INTERVention disinfecting wipes = 'disinfectant wipes'
- Accel 5 TB disinfectant (liquid) = 'disinfectant (liquid)'
- Alcohol based hand rub = 'ABHR'
- Blue biohazardous waste container = 'blue bin'

1.	In AFB room, doff PPE with assistance of the buddy technologist or a Trained Observer, following doffing protocol: <i>'Donning and Doffing Viral Hemorrhagic Fever (VHF) Specimen Processing PPE Check List for VGH Laboratory Technologists'</i> ; Buddy technologist or Trained Observer reading checklist and observing from outside AFB Room. Place all PPE into blue bin. Do not press down to compress contents of bin.
2.	Put on new gloves and a routine lab gown.
3.	Tie off red bags with twist ties, one inside the other, taking care to avoid contact with gown or self to the blue bin.

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4.	Disinfect gloves with disinfectant wipes or alcohol-based hand rub (ABHR). Allow to dry.
5.	Using the supplied silver marker, write the "Quarantine Patient #" (as issued by Infection Control) on the label provided. Place label on outside of blue bin.
6.	Place two biohazard symbols on the blue bin so that they are visible from two sides.
7.	<p>If the patient's status is positive for Ebola VD/VHF, also affix the following provided labels so that they are visible from two sides of the drum:</p> <ul style="list-style-type: none"> • Two 'Infectious Substance Affecting Humans UN2814 Class 6.2' • Two 'Incinerate Only'
8.	Place lid on bin. Crothall will clamp lid upon removal.
9.	Remove routine gown if used and place in box provided for used AFB processing gowns.
10.	Disinfect gloves with disinfectant wipes or ABHR. Allow to dry. Using glove to glove technique, remove gloves and place in routine biohazard waste.
11.	Wipe surface of Accel INTERvention wipes container and the ABHR container with disinfectant wipes.
12.	Carefully perform hand hygiene with ABHR or with soap and water at sink.
13.	Leave AFB room, close door and secure room leaving signage in place and applying barrier/masking tape over the door to prevent accidental entry.
14.	Proceed according to the patient's current status:
	<p>a. If the patient's Ebola VD/VHF status is still pending:</p> <p>- Call Crothall at 1-844-372-1959 and request: "Blue Bin removal for Suspect Ebola VD/VHF", "STAT specialized terminal clean for Ebola VD/VHF"</p>
	<p>b. If the patient's Ebola VD/VHF status is positive:</p> <p>- Call Crothall at 1-844-372-1959. Place request for: "Blue Bin for Positive Ebola VD/VHF", "STAT specialized terminal clean for Ebola VD/VHF"</p>
15.	AFB room cannot be used until Crothall removes all waste and completes the final room cleaning.

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J: Day 2/ Secondary Inactivated Plates:

- If patient's status is pending, check with medical microbiologist for any updates before proceeding.
- Ask the Medical Microbiologist to sign 'Waste Release Form'.
 - Note: Accel INTERvention disinfecting wipes require one minute contact time
Accel 5 TB disinfectant (liquid) requires five minute contact time
 - Accel INTERvention disinfecting wipes = 'disinfectant wipes'
 - Accel 5 TB disinfectant (liquid) = 'disinfectant (liquid)'
 - Alcohol based hand rub = 'ABHR'
 - Blue biohazardous waste container = 'blue bin'

v		Step
	1.	AFB technologist will have the buddy technologist or Trained Observer to assist them in donning and doffing following the lab protocol checklist: <i>'Donning and Doffing Viral Hemorrhagic Fever (VHF) Specimen Processing PPE Check List for VGH Laboratory Technologists'</i> . Following full PPE donning, the AFB technologist will perform the Day 2 work up independently.
	2.	Sign <i>Specimen Handling Contact List</i> for specimen processing.
	3.	Place absorbent material provided at the bottom of the Blue bin; double line the bin with red bags and place in AFB room.
	4.	Place sign on AFB room door 'Suspect Ebola VD/VHF Processing and Containment. DO NOT ENTER'.
	5.	Remove secondary plates from incubator and place into BSC. Confirm that all plates are labelled with an 'inactivated' sticker.
	6.	Wipe outside of plates with disinfectant wipes.
	7.	Check to confirm growth on secondary plates. Repeat subculture from primary plates if necessary following the pm growth protocol. Remove inactivated plates from BSC.
	8.	Wipe all surfaces inside the BSC and all surfaces used outside the BSC using Accel Five TB. Allow to remain wet for minimum 5 minutes.
	9.	Wipe all the exterior surfaces of the blue bin with disinfectant wipes, then wipe the exterior surfaces of the disinfectant containers
	10.	Place an absorbent mat on the floor and saturate with disinfectant (liquid). Place the blue bin on top of the mat.

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	11.	Remove PPE in AFB room with the assistance of the buddy technologist or Trained Observer, following doffing protocol checklist; Buddy technologist or Trained Observer reading checklist and observing from outside AFB Room. Place all PPE into blue bin. Do NOT compress contents of blue bin.
	12.	Give secondary plates to the BC technologist. Secondary inactivated plates can be worked up on BC bench per usual procedures and using usual laboratory precautions. All 'inactivated' labelled plates can be discarded per routine protocol.

K: Waste removal from AFB room:

- Follow the waste removal protocol found in Section I

L: Day 3/ Examination of Anaerobic Plates

- If patient's status is pending, check with medical microbiologist for any updates before proceeding
- Ask the Medical Microbiologist to sign 'Waste Release Form'

Follow protocol as for G: Subculture of primary plates to secondary:

- Prepare EVD/VHF positive blood culture supply box with all required materials, including a new anaerobic jar and media plates for subculture of anaerobic organisms on primary plates to secondary plates.
- Include the protocols in sections G and H and the donning/doffing protocol: '*Donning and Doffing Viral Hemorrhagic Fever (VHF) Specimen Processing PPE Check List for VGH Laboratory Technologists*', disposable loops, blue absorbent pads, a double lined cardboard bucket for waste.
- Consider the gram stain results and the presence of any aerobic growth, allow for the possibility of more than one colony type being present.
- AFB (or designated) technologist will perform the work up independently but will have the buddy technologist or Trained Observer for the donning and doffing following the lab doffing checklist.

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V		Step
	1.	Take all necessary supplies to the AFB Room.
	2.	Place down blue absorbent pads and arrange all materials inside hood according to safe practice use of a BSC. Refer to instructions for proper use of BSC – laminated copy posted near AFB BSC.
	3.	Sign the Specimen Handling Infection Prevention sheet.
	4.	Don PPE with buddy technologist or Trained Observer.
	5.	Place sign on door to AFB Room: “Suspect Ebola VD/VHF Containment and Processing. DO NOT ENTER”. Close AFB Room door.
	6.	Place and open the anaerobic jar in the BSC and examine plates.
	7.	If anaerobic growth is suspected, follow the protocol in G for the subculture of primary plates to secondary plates.
	8.	Incubate secondary plates in a new anaerobic jar labelled with ‘inactivated’ sticker.
	9.	<p>Disinfect original anaerobic jar using disinfectant wipes and place into red biohazardous bags (double bagged); tie off one bag inside the other. Label using a white label with red edging and bold lettering: “Ebola VD/VHF status pending. Do not remove from AFB Room”</p> <p>This will be stored in BSC until the final status of patient is determined. If patient tests EVD/VHF negative, jar will be put back into regular use. If patient tests EVD/VHF positive, jar will be discarded into Blue bin.</p>
	10.	Proceed with AFB Room Cleanup as described in Section H and in Waste Removal as in Section I

Subsequent work should follow the protocol listed in section J: Secondary Plates, as well as the cleanup and waste removal as described in Sections H and I respectively.

